



**BIOL 3702L:
Smear
Preparation
and Simple
Staining**

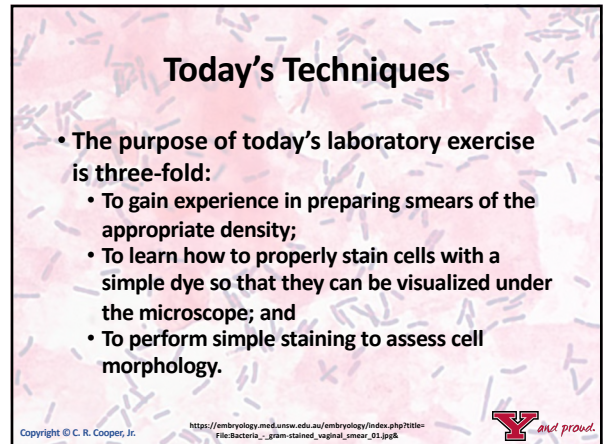
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<http://www.aviano.af.mil/News/Article-Display/Article/724769/mda.girmen-ensure-success-through-support/>

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Today's Techniques

- The purpose of today's laboratory exercise is three-fold:
 - To gain experience in preparing smears of the appropriate density;
 - To learn how to properly stain cells with a simple dye so that they can be visualized under the microscope; and
 - To perform simple staining to assess cell morphology.




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https://embryology.med.unsw.edu.au/embryology/index.php?title=File:Bacteria_-_gram_stained_vaginal_smear_01.jpg&

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Smear Preparation

- Making a heat-fixed smear is more of an "art" than it is science.
- The goal is to make a smear that is dense enough, but not too dense, to permit both cell visualization and proper staining
 - Too dense – hard to distinguish cells and staining patterns can be affected. **AVOID MAKING DENSE SMEARS!!!**
 - Too sparse – difficult to find cells under the microscope




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Three Smears of Differing Thickness
(marked by white wax ring)

- Left and right smears: **too thick**
- Center smear: **correct thickness**




<https://hicksmicro.blogspot.com/p/staining-problems.html>

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Smear Preparation (cont.)

- Two types of cultures can be used to make smears and each requires slightly different approaches
 - **Solid cultures** – aseptic transfer of a very **small amount** of cells to a **small volume** of water on the surface of a microscope slide
 - **Broth cultures** – aseptic transfer of a **one or more drops** of culture fluid spread out on the surface of a microscope slide



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Making a Bacterial Smear – Solid Culture



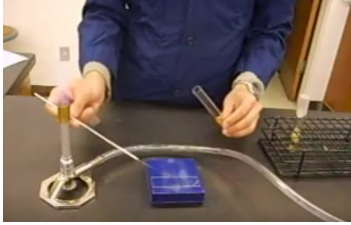
https://youtu.be/gM5_Y2QgXq0

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Making a Bacterial Smear – Broth Culture



<https://youtu.be/HxiN2mBLGCE>

What obvious error is being committed during this procedure?

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Smear Preparation (cont.)

- Following the preparation of the smear and allowing it to air dry, the cells must be heat-fixed by one of the following methods:
 - **(PREFERRED METHOD)** Pass the slide through the top blue portion of the Bunsen burner flame – three to six passes at no more than one second each
 - Place on a hot plate set to the highest temperature for no more than three seconds

USE EXTREME CAUTION WITH HEAT SOURCES!!!

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Smear Preparation (cont.)

- Once heat-fixed, the smears are now ready for staining
- **STAINING MUST TAKE PLACE ONLY ON THE PROVIDED RACKS SET OVER THE BENCH SINKS!!!** (This is one of the few times that chemical reagents are permitted to flow into the sinks.)

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Simple Staining of a Bacterial Smear



<https://youtu.be/n5fXlpJUGD4>

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Staining Slides

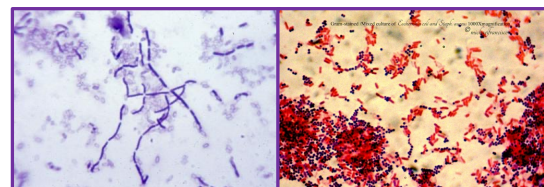
- Stains can be used not only to visualize cells and distinguish cell shape, but with the appropriate staining procedure (e.g., Gram stain), distinct cell types can be observed.
- Simple staining involves a single dye placed on a smear, then washed away prior to observation under a microscope.
- The types of dyes and the length of staining times differ with a given procedure.

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Staining Slides (cont.)



Simple stain (left) and Gram stain (right)

<https://www.studyblue.com/notes/n/microbio-lab-practical/deck/3837183>
and <https://www.flickr.com/photos/micodude/6500465759>

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Today's Exercise

- **Smears from solid cultures:**
 - On two (2) separate slides, place a small drop of water from the bench-top water bottle. Use these slides to prepare a single smear each of
 - *Staphylococcus aureus* from a TSA slant culture
 - *Escherichia coli* from a TSA slant culture
 - Allow the slides to dry, then heat fix both slides.

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Today's Exercise (cont.)

- **Smears from a broth culture:**
 - Completely suspend the cells from the TSB mixed culture of *Escherichia coli* and *Staphylococcus aureus* by rolling the tube between your hands
 - Prepare a smear from the TSB mixed culture.
 - Allow to dry, then heat fix the slides.

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Today's Exercise (cont.)

- **Staining with Crystal Violet (CV):**
 - Place the heat-fixed slides on the staining rack
 - Flood the area of the smear on each slide with CV; allow the CV to remain on the slides for 60 to 90 seconds
 - Drain the CV from the slides and briefly wash (3-4 seconds) the slide with water from the bench-top bottle.

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Today's Exercise (cont.)

- **Staining with Crystal Violet (CV): (cont.)**
 - Drain the water from the slides and carefully blot the slides dry using **PAPER TOWEL**. (Bibulous paper is not available.)
 - Examine the slides under oil immersion and record your observations on the exercise data sheet. **NO COVERSLIP IS NEEDED!!!!**

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Today's Exercise (cont.)

- **Special Notes/Hints:**
 - 1) All staining can be done at the same time.
 - 2) Slides that are heat-fixed or stained can be stored in slide boxes for later processing.
 - 3) To make a smear from a broth culture, the number of loopfuls is dependent upon the density (turbidity) of the culture.

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Today's Exercise (cont.)

- **Special Notes/Hints: (cont.)**
 - 4) Broth cultures that have sat for a time should be thoroughly mixed by rolling the tube between your palms or sharply thumping the tube several times.
 - 5) No coverslips are needed on fixed, stained smears. To observe cells, place the immersion oil directly on the smear.

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Lab Report Expectations

- Be sure to complete laboratory worksheet.
- Retain this lab report. Your laboratory instructor may call for its submission in the near future.

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